The phylogenetic signal in cranial morphology of *Vipera aspis*: a contribution from geometric morphometrics

A. Gentilli¹, A. Cardini², D. Fontaneto³ & M.A.L. Zuffi⁴

¹Dipartimento di Biologia Animale, Università di Pavia, Italy ²Dipartimento del Museo di Paleobiologia e dell'Orto Botanico, Università di Modena e Reggio Emilia, Italy & Hull York Medical School, The University of Hull, UK ³Imperial College London, Division of Biology, UK ⁴Museo di Storia Naturale e del Territorio, Università di Pisa, Italy

Morphological variation in the frontal bone and cranial base of *Vipera aspis* was studied using geometric morphometrics. Significant differences in shape were found among samples from subspecies present in Italy (*V. a. aspis*, *V. a. francisciredi*, *V. a. hugyi*). Sexual dimorphism was negligible as well as allometry and size differences. The most divergent subspecies was *V. a. aspis*, possibly in relation to its recent history of geographic isolation in a glacial refugium. Shape clusters were in good agreement with clusters from studies of external morphology and completely congruent with results from molecular studies of mtDNA.

Key words: asp viper, cranial shape and size, Italy, phylogeny, systematics

INTRODUCTION

he asp viper, Vipera aspis (Linnaeus, 1758), is among the best studied Palaearctic viperids (Mallow et al., 2003). It is a small to medium-sized snake, averaging 60-70 cm total length (50-63 cm snout-vent length), up to 82 cm maximum length (Saint Girons, 1978). It is only found in western and southern Europe (Zuffi, 2002; Fig. 1) from sea level to 3000 m (Saint Girons, 1997). Geographic variability in morphological features has been described (Phisalix, 1968; Kramer, 1980; Zuffi & Bonnet, 1999; Zuffi, 2002). For instance, Zuffi (2002) studied characters of the external morphology and copulatory apparatus, and found differences in the number of dorsal bars and of ventral and subcaudal scales, together with a distinctive hemipenial form. Thus, he suggested that Vipera aspis may indeed represent a species complex (sensu Hermann et al., 1999) including Vipera aspis, V. atra Meisner 1820, V. hugyi Schinz 1833 and V. zinnikeri Kramer 1958. A taxonomic revision was also carried out using immunological data by Pozio (1980), who showed that electrophoretic patterns of venom compositions were very different between V. aspis atra, V. aspis francisciredi Laurenti, 1768, and V. aspis hugyi, whereas the latter had the same electrophoretic pattern as V. aspis montecristi Mertens, 1956. However, the reproductive isolation of populations within the V. aspis species group has been questioned by some authors focusing on neurotoxins. For instance, Guillemin et al. (2003) found that interbreeding may have occurred even between two clearly separated species, V. aspis from southeastern France and the sand viper, Vipera ammodytes (Linnaeus 1758). On the other hand, De Haro et al. (2002) showed that in France at least one population of V. aspis had a specific venom profile distinct from

those of other French populations. Garrigues et al. (2005) also made a preliminary comparison using mitochondrial cytochrome b and ND2 DNA sequences. However, the limited number of samples analysed by Garrigues et al. (2005) did not allow any robust reconstruction of relationships within V. aspis. In contrast, a recent study on mtDNA variability investigating the phylogeography of the asp viper across its whole distribution range (Ursenbacher et al., 2006) indicated that V. aspis can actually be separated into four distinct groups. Moreover, they considered V. atra, characterized by a relatively high number of ventral scales and a marked black dorsal pattern, and distributed in northwestern Italy (Zuffi, 2002), as a synonym of V. a. aspis. Thus, the four well-supported molecular clades, corresponding to V. a. aspis, V. a. francisciredi, V. a. hugyi and V. a. zinnikeri, were considered as subspecies.

Whether populations that are recognized using softtissue anatomy and molecular markers can also be discriminated using bone morphology has never been accurately investigated. Moreover, incongruences between results from previous studies are present, requiring further analyses and the clarification of independent characters.

Cranial form provides potentially interesting traits for investigating the systematic relationships of vipers (Guo & Zhao, 2006). In particular, the akinetic portion of the skull has been considered more suitable for taxonomic comparisons in snakes (Kramer, 1980; Gloyd & Conant, 1990). This is because the neurocranium, which forms the akinetic portion of the skull, is less strongly influenced by selective pressures related to diet (prey swallowing) and it is not directly associated with the mechanics of envenomation. Such studies, however, either did not focus on

Correspondence: M.A.L. Zuffi, Museo di Storia Naturale e del Territorio, Università di Pisa, via Roma 79, 56011 Calci (Pisa), Italy. *E-mail*: marcoz@museo.unipi.it

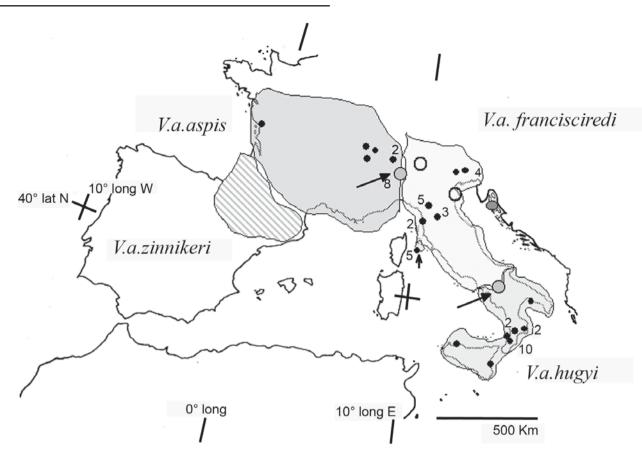


Fig. 1. Schematic distribution of *Vipera aspis* group (from Zuffi, 2002, modified, according to Ursenbacher et al., 2006). Small black dots indicate sampling localities of *V. aspis* subspecies (numbers indicate sample size >1); two large open dots indicate *V. berus* outgroups; the large dark grey dot indicates *V. ammodytes* outgroup. The small arrow at the black dot indicates the *V. a. hugyi* population of Montecristo Island. The large arrows at large grey dots indicate the transition forms, *V. a. aspis*francisciredi* in the north, and *V. a. francisciredi*hugyi* in the south (see Appendix 1 for references).

European vipers or were limited by the paucity of available information in terms of study specimens and number of metric characters. Morphometric tools are presently available that allow accurate comparisons of the geometric shape of organisms or of their organs (Rohlf & Marcus, 1993; Adams et al., 2004). These methods, known as geometric morphometrics, have been successfully applied to detect subtle variation among closely related taxa (Duarte et al., 2000; Cardini, 2003; Frost et al., 2003; Klingenberg et al., 2003; Nicola et al., 2003; Macholán, 2006).

Thus, we applied geometric morphometrics to explore size and shape variation in the cranium of Italian populations of *V. aspis* that include three of the four subspecies supported by the molecular analysis of Ursenbacher et al. (2006). The analysis was performed by measuring crania using two-dimensional coordinates of anatomical landmarks in representatives of *V. a. aspis*, *V. a. francisciredi* and *V. a. hugyi*. The main aims of the study were to assess the magnitude of the variation and the significance of differences in cranial size and shape among these subspecies, and to reconstruct their similarity relationships.

MATERIALS AND METHODS

We considered only adult specimens (*n*=61) recognized and classified following Zuffi & Bonnet (1999). *Vipera a. aspis* (4 males, 3 females), *V. a. francisciredi* (10 males, 8 females) and *V. a. hugyi* (14 males, 9 females) (*sensu* Ursenbacher et al., 2006) were analysed. Individuals from contact areas between species, also showing intermediate numbers of dorsal bars and markings and numbers of ventral scales, were considered separately, and indicated as *V. a. aspis*francisciredi* (5 males, 3 females) and *V. a. francisciredi*hugyi* (1 male). Specimens of *V. berus* (1 male, 2 females) and *V. ammodytes* (1 female), closely related to *V. aspis* (Hermann et al., 1999), were used as outgroups. Museum catalogue numbers and localities of collection are given in Appendix 1.

Digital pictures of skulls were used in the analysis. Pictures of the dorsal and ventral view of the akinetic portion of the cranium were taken in standardized conditions using a Leica Digilux Zoom digital photocamera, with a 300 dpi resolution, mounted on a Leica MZ75 stereomicroscope, and lit by optical fibres. A set of topographically corresponding anatomical landmarks (Marcus et al., 2000)

	Description
Dorsal	
1	Proximal margin of the suture between the frontals
2	Outermost point between prefrontals and frontals
3	Point between the frontal and postfrontal bones
4	Suture point between the frontal, postfrontal and parietal bones
5	Suture point between frontals and parietals
Ventral	
1-2-3-4	Higher points of the basisphenoid lateral crest
5	Teeth of the basioccipital

on the frontal bone and cranial base were used as morphometric descriptors (Fig. 2, Table 1). Information on only one side of the cranium was used in order to avoid redundancy in symmetric structures. Five almost coplanar and clearly visible landmarks were used for describing the shape of the frontal bone (Fig. 2A). Similarly, five approximately coplanar landmarks were used to describe the shape of the basisphenoid and basioccipital bones in the cranial base (Fig. 2B). Crania were not always well preserved and either ventral or dorsal bones were missing in a minority of specimens. For these, either only the frontal bone or the cranial base was therefore measured.

We applied geometric morphometrics techniques (Bookstein, 1991; Dryden and Mardia, 1998; Zelditch et al., 2004), implementing linear statistical models (Rohlf, 1998; Klingenberg & Monteiro, 2005). Differences in landmark coordinates due to position or size were removed with a generalized procrustes analysis (GPA – Rohlf & Slice, 1990), thus leaving only information on shape in the landmark configurations. Size was computed as centroid size (CS), which is a measure of the dispersion of landmarks around the barycentre of the configuration. Digitizing error and tangent space approximation to the space shape, estimated as in Cardini & Tongiorgi (2003), were found not to introduce any appreciable error.

Differences between groups were described with thin plate spline (TPS) deformation grids (Bookstein, 1991; Adams et al., 2004). After testing for differences in shape of frontal and ventral bones among groups (sex × species MANOVA for *V.a. aspis*, *V. a. francisciredi*, *V. a. hugyi*, and *V. a. aspis*francisciredi*), the two data sets were combined by appending the matrices of frontal and cranial base shape variables (Adams, 1999), and MANOVA (sex × species) and CVA (pooled sexes) were performed to test the significance of group differences using all available information. A scatterplot of the principal components of the shape was used to show graphically the relationships among individuals and groups. The relationship between size and shape in adults (static allometry) was assessed in the two largest samples (*V. a.* francisciredi and V. a. hugyi) by regressing shape variables onto CS. Significance of regressions was tested with a permutation test for the generalized Goodall's F (Goodall, 1991; Rohlf, 2005). Average shapes for the frontal and cranial base were computed for each species with pooled sexes. The two data sets were then combined (Adams, 1999) and phenetic relationships among viper taxa were summarized with multi-dimensional scaling (MDS) and cluster analysis (performed on the matrix of Euclidean distances between mean shapes). A minimumlength spanning tree (MST; Rohlf, 1970) was superimposed on the MDS scatterplot to help detect local distortions. The matrix correlation (a Pearson correlation on unfolded diagonal symmetric matrices) between the original matrix of Euclidean distances (all shape variables) and the one based on the position of the species in the three-dimensional MDS scatterplot was 0.960. Among different clustering methods, the unweighted pair-group method using centroid (UPGMC) had the highest cophenetic correlation (Rohlf, 1970) to the original distance matrix (r=0.856); thus, UPGMC was used for the cluster analysis. All analyses were performed in SPSS and NTSYSpc 2.1.

To quantify the congruence between shape similarity relationships and phylogeny, we computed a matrix correlation between patristic distances from the UPGMC phenogram and those from the gene tree of Ursenbacher et al. (2006) as proxy for phylogeny based on independent data. The phylogenetic tree was built using different haplotypes for each species; for the comparison, we selected haplotypes by choosing individuals from localities closest to our study samples, i.e. haplotype H1 for *V. a. aspis*, H16 for *V. a. francisciredi* and H20 for *V. a. hugyi* (Ursenbacher et al., 2006).

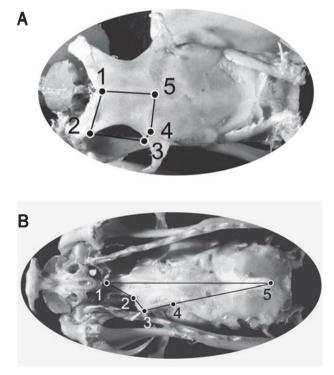


Fig. 2. Landmark configurations for frontal bone (A) and cranial base (B) of the vipers.

View	Effect	$\boldsymbol{\lambda}_{_{Wilks}}$	F	d.f.	Р
Frontal	Sex	0.792	1.663	6,38	0.157
	Group	0.386	2.403	18, 108	0.003**
	$\operatorname{Sex} \times \operatorname{group}$	0.580	1.274	18, 108	0.219
Cranial base	Sex	0.770	1.097	6,22	0.395
	Group	0.246	2.234	18,62.7	0.010*
	$\operatorname{Sex} \times \operatorname{group}$	0.654	0.565	18,62.7	0.911
Combined	Sex	0.028	1.401	12, 14	0.271
	Group	0.454	2.733	36,42	0.001**
	$\operatorname{Sex} \times \operatorname{group}$	0.232	0.748	36,42	0.811

Table 2. MANOVA sex × species of shape variables for frontal and cranial base of *V. a. francisciredi*, *V. a. aspis*, *V. a hugyi* and *V. a. aspis* * *francisciredi* and for the combined data set. Significant values: *P<0.05; **P<0.01.

RESULTS

Separate analyses of frontal bone and cranial base

Size differences (results not shown) between groups were generally negligible in both datasets (frontal bone and cranial base). Only V. a. francisciredi and V. a. hugyi showed appreciable though very small differences in frontal bone size (the former being the largest species in the group). Size was also generally homogeneous in each group as suggested by coefficients of variation (ratio between SD and mean) ranging between 5.2% and 15.8%. Group differences were significant for shape of both frontal bone and cranial base of V. a. aspis, V. a. francisciredi, V. a. hugyi and V. a. aspis*francisciredi, while sexual dimorphism and the interaction term were not (Table 2). The same outcome was found in the combined data set (Table 2). Sexual dimorphism was negligible compared to group differences. Thus, groups were compared with pooled sexes in further analyses.

Static allometry was investigated in *V. a. francisciredi* and *V. a. hugyi*. The proportion of shape variation corre-

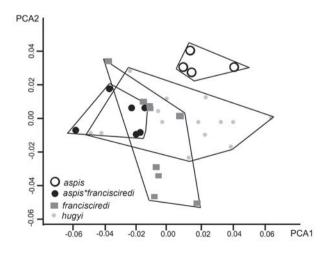


Fig. 3. Species discrimination in the *V. aspis* group using the combined data set (frontal bone + cranial base). First two axes of a PCA of all shape variables. PCA1 explains 44.2%; PCA2 explains 21.1%.

lated to size was small (7.2% and 6.1% respectively) and not significant. Similar results were found when sexes were analysed separately.

Combined dataset

Differences among V. a. francisciredi, V. a. aspis, V. a. hugyi and V. a. aspis*francisciredi cranial shapes were significant (CVA, l_{wilks} =0.0430, $F_{36,53}$ =2.848, P<0.001, total hit ratio 90.9%). Tests of pairwise differences based on Mahalanobis distances were significant, except for the comparison between V. a. francisciredi and V. a. aspis*francisciredi (Table 3). Also, V. a. francisciredi and V. a. aspis*francisciredi (Table 3). Also, V. a. francisciredi and V. a. hugyi are relatively similar (P>0.05). Group separation in the scatterplot of the specimens along the first two PCA axes (Fig. 3) suggests that Vipera a. aspis is most distinctive.

The MDS scatterplot and UPGMC phenogram of mean shapes suggest phenetic relationships congruent with those presented above (Figs. 4-5). Vipera a. francisciredi*hugyi is close to V. a. hugyi; both are characterized by medially elongated frontal bones with a relatively narrow posterior region. This region is comparatively larger in V. a. francisciredi and V. a. aspis*francisciredi, whose strong similarity are suggested by both the ordination and the phenogram. As in the PCA of individual specimens (Fig. 3), V. a. aspis is distinctive, especially in the cranial base, and intermediate between the outgroups and all other subspecies. Finally, as expected, outgroup species (V. ammodytes and V. berus), with their small anterior basisphenoid (landmarks 1-4) and elongated posterior cranial base (Fig. 5), are distinct from V. aspis.

Patristic distances of *V. a. francisciredi*, *V. a. aspis* and *V. a. hugyi* from the UPGMC phenogram are strongly in agreement with those from the phylogenetic tree of Ursenbacher et al. (2006) (r=0.992).

DISCUSSION

Tests of sexual dimorphism were performed to decide whether pooling samples regardless of sex was warranted. Indeed, sexual dimorphism was not evident in our analysis and this is consistent with results from ecological studies that found no dietary differences between female and male vipers (Luiselli & Agrimi, 1991). Besides, selective pressures that might increase sexual dimorphism may be moderate in this species as bites are not

,	8			
	V. a. francisciredi	V. a. aspis	V. a. hugyi	V. a. aspis*francisciredi
V. a. francisciredi		37.409	9.306	6.588
V. a. aspis	0.003**		31.156	36.560
V. a. hugyi	0.047*	0.005**		18.333
V. a. aspis*francisciredi	0.451	0.008**	0.009**	

Table 3. Test of pairwise differences in shape from the combined data set. Significance levels (*P<0.05; ** P<0.01) are below the main diagonal and Mahalanobis squared distances above.

used during courtship and combat by males (Andrén, 1986).

The main focus of the study was to assess whether subspecies distinguished on the basis of external morphology (Zuffi & Bonnet, 1999; Zuffi, 2002; Zuffi et al., 2003), and supported by a recent molecular analysis (Ursenbacher et al., 2006), were also supported by variation in cranial size and shape. Indeed, shape was significantly different and relationships among Italian subspecies of V. aspis were completely congruent with molecular clades (Ursenbacher et al., 2006). Vipera a. francisciredi and V. a. hugyi were more similar to each other than any were to V. a. aspis, as in the mtDNA phylogenetic tree by Ursenbacher et al. (2006). Moreover, even among subspecies the relationships identified by the analysis of cranial shape were congruent with results from mtDNA phylogeny. For example, the population from Montecristo Island, traditionally believed a valid subspecific taxon (V. a. montecristi), had the same cranial shape as the specimens of V. a. hugyi we analysed from southern Italy. We thus confirmed that the population from Montecristo Island may be considered a recent introduction from populations of V. a. hugyi (Zuffi, 2002; see also Barbanera et al., 2009), as also previously suggested by its venom composition (Pozio, 1980). The monotypy of V. a. aspis was also confirmed: populations

from the western Alps had the same cranial shape as those from northern France, as expected from results on mtDNA phylogeny, and from external morphological analyses of scale and dorsal pattern variation (Golay et al., 2008).

Size, in contrast, did not show appreciable group differences. Cranial size might be evolutionarily more labile than shape and thus often unable to preserve a strong phylogenetic signal. This might be explained by the higher complexity of shape. Oxnard (2000) observed that morphometric comparisons of individual skeletal units (such as arms, limbs or teeth) tend to produce clusters that indicate functional convergences of anatomical parts in primates. In contrast, when variables from different anatomical regions are combined in a single analysis, separations of species mostly reflect evolutionary relatedness. He suggested that phylogenetic information within a structure is relatively small, whereas when several structures are analysed together, phylogenetic groups emerge (Oxnard, 2000). Similarly, the informativeness of a multivariate morphological descriptor like shape might be much larger than that of univariate size and thus more likely to pick up differences of phylogenetic interest. That shape might provide insight into the evolutionary history of this group was also suggested by the observation that V. a. aspis had the

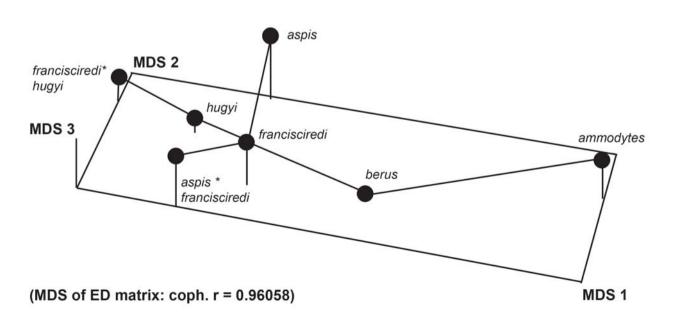


Fig. 4. Phenetic relationships among *Vipera* species (including the outgroups and the two transition forms). MDS scatterplot of the mean shapes.

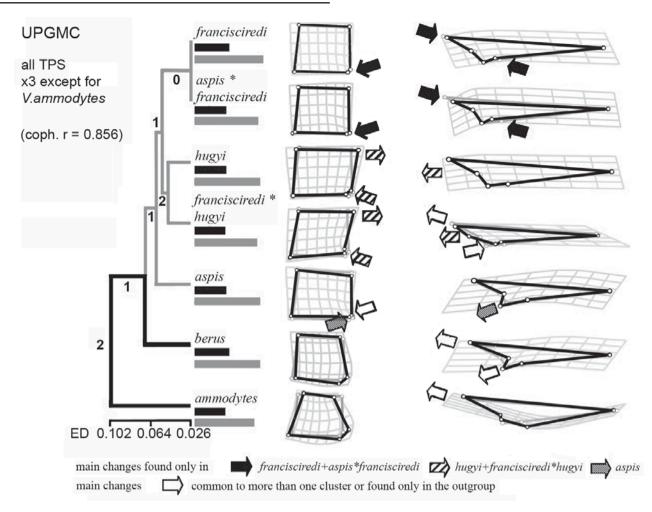


Fig. 5. UPGMC phenogram of the mean shapes using the combined data set. TPS deformation grids for the frontal and cranial base are shown by each taxon: they are magnified (cranial base of *V. ammodytes* × 1.5; other taxa × 3) to emphasize shape differences between a taxon (black) and the mean of all taxa (grey). Arrows help detect shape features typical of different taxa. Lines below species names are proportional to the average centroid size of that taxon (black for the dorsal view and grey for the ventral view). Numbers by each branch indicate whether that cluster is found in both 50% subsets (2), in just one of them (1) or in none of them (0).

most distinctive cranial shape. The geographic range of this subspecies in southeastern France and northwestern Italy overlaps with an area considered an important glacial refugium of the asp viper (Ursenbacher et al., 2006). Here, geographic isolation in a period of environmental change might have promoted morphological divergence.

In summary, the samples of *V. aspis* studied were significantly different, but clustered together to the exclusion of outgroup species. This was again congruent with the conclusion of Ursenbacher et al. (2006) that taxa within this group show differences smaller than those between well recognized separate species of *Vipera*.

Our samples also included two taxa that could not be unequivocally ascribed to defined subspecies, but had taxonomic features intermediate between two subspecies. One of them, V. a. francisciredi*hugyi, was very close to V. a. hugyi, while the other, V. a. aspis*francisciredi, was very similar to V. a. francisciredi. We cannot support or refute the hypothesis of a supposed "hybrid" origin of these populations due to poor sampling; nevertheless, notwithstanding intermediate taxonomic features, cranial shape was very similar to only one of the supposed parental taxa. Larger samples will be needed to accurately quantify and compare cranial variation in *V. aspis* and carefully compare shape and size of individuals from contact areas. Indeed, the present study is aimed at providing a preliminary investigation of differences in *V. aspis* using hard-tissue morphology. The paucity of specimens from this group in museum collections strongly limited the size of our samples. This is a common problem in taxonomic analyses (Marcus, 1990), where preliminary investigations often help to stimulate extensive follow-up studies.

ACKNOWLEDGEMENTS

We wish to thank M.D. Candia Carnevali (Department of Biology, University of Milan) for the use of her laboratory's optical and digital instruments, and Marta Poggesi and Annamaria Nistri for providing access to the Enrica Calabresi herpetological collection (Zoological Museum, University of Florence).

REFERENCES

- Adams, D.C. (1999). Methods for shape analysis of landmark data from articulated structures. *Evolutionary Ecological Research* 1, 959–970.
- Adams, D.C., Rohlf, F.J. & Slice, D.E. (2004). Geometric morphometrics: ten years of progress following the 'revolution'. *Italian Journal of Zoology* 71, 5–16.
- Andrén, C. (1986). Courtship, mating and agonistic behaviour in a free-living population of adders, *Vipera berus* (L.). *Amphibia–Reptilia* 7, 353–383.
- Barbanera, F., Zuffi, M.A.L., Guerrini, M., Gentilli, A., Tofanelli, S., Fasola, M. & Dini, F. (2009).
 Phylogeography of the Italian asp viper (*Vipera aspis*) as inferred from mitochondrial and microsatellite DNA data. <u>Molecular Phylogenetics and Evolution 52, 103–</u> 114.
- Bookstein, F.L. (1991). *Morphometric Tools for Landmark Data*. Cambridge: Cambridge University Press.
- Calabresi, E. (1924). Ricerche sulle variazioni della Vipera aspis Auct. in Italia. Bollettino Istituto di Zoologia della Reale Università di Roma 2, 78–127.
- Cardini, A. (2003). The geometry of marmot (Rodentia: Sciuridae) mandible: phylogeny and patterns of morphological evolution. <u>Systematic Biology</u> 52, 186– 205.
- Cardini, A. & Tongiorgi, P. (2003) Yellow-bellied marmots 'in the shape space': sexual dimorphism, growth and allometry of the mandible. <u>Zoomorphology</u> 122, 11–23.
- De Haro, L., Robbe-Vincent, A., Saliou, B., Valli, M., Bon, C. & Choumet, V. (2002). Unusual neurotoxic envenomations by *Vipera aspis aspis snakes* in France. *Human and Experimental Toxicology* 21, 137–145.
- Dryden, I.L. & Mardia, K.V. (1998). *Statistical Shape Analysis*. New York: John Wiley and Sons.
- Duarte, L.C., Monteiro, L.R., Von Zuben, F.J. & Dos Reis, S.F. (2000). Variation in the mandible shape in *Thrichomys apereoides* (Mammalia: Rodentia): geometric analysis of a complex morphological structure. *Systematic Biology* 49, 563–578.
- Frost, S.R., Marcus, L.F., Bookstein, F.L., Reddy, D.P. & Delson, E. (2003). Cranial allometry, phylogeography, and systematics of large-bodied papionins (Primates: Cercopithecinae) inferred from geometric morphometric analysis of landmark data. *Anatomical Record* 275A, 1048–1072.
- Garrigues, T., Dauga, C., Ferquel, E., Choumet, V. & Failloux, A.-B. (2005). Molecular phylogeny of Vipera Laurenti, 1768 and the related genera Macrovipera (Reuss, 1927) and Daboia (Gray, 1842), with comments about neurotoxic Vipera aspis aspis populations. Molecular Phylogenetics and Evolution 35, 35–47.
- Gloyd, H.-K. & Conant, R. (1990). Snakes of the Agkistrodon Complex. A Monographic Review. Oxford, Ohio: Society for the Study of Amphibians and Reptiles.
- Golay, P., Monney, J.-C., Monelli, A., Durand, T., Thiery,G., Zuffi, M.A.L. & Ursenbacher, S. (2008).Systematics of the Swiss asp viper: some implications

for the European *Vipera aspis* (Linnaeus 1758) complex (Serpentes: Viperidae). A tribute to Eugen Kramer. *Amphibia–Reptilia* 29, 71–83.

- Goodall, C.R. (1991). Procrustes methods in the statistical analysis of shape. *Journal of the Royal Statistical Society* B53, 285–339.
- Guillemin, I., Bouchier, C., Garrigues, T., Wisner, A. & Choumet, V. (2003). Sequences and structural organization of phospholipase A2 genes from Vipera aspis aspis, V. aspis zinnikeri and Vipera berus berus venom. Identification of the origin of a new viper population based on ammodytin I1 heterogeneity. European Journal of Biochemistry 270, 2697–2706.
- Guo, P. & Zhao, E.M. (2006). Comparison of skull morphology in nine Asian pit vipers (Serpentes: Crotalinae). *Herpetological Journal* 16, 305–313.
- Herrmann, H.-W., Joger, U., Lenk, P. & Wink, M. (1999).
 Morphological and molecular phylogenies of viperines: conflicting evidence? In *Phylogeny and Systematics of the Viperidae, Kaupia, Vol.* 8, 21–30. Joger, U. (ed.).
 Darmstadt: Hessisches Landesmuseum.
- Klingenberg, C.P. & Monteiro, L.R. (2005). Distances and directions in multidimensional shape spaces: implications for morphometric applications. <u>Systematic</u> *Biology* 54, 678–688.
- Klingenberg, C.P., Barluenga, M. & Meyer, A. (2003). Body shape variation in cichlid fishes of the Amphilophus citrinellus species complex. <u>Biological Journal of the</u> Linnean Society 80, 397–408.
- Kramer, E. (1980). Zum Skelett der Aspisviper, Vipera aspis (Linnaeus, 1758). Revue Suisse de Zoologie 87, 3– 16.
- Luiselli, L. & Agrimi, L. (1991). Composition and variation of the diet of *Vipera aspis francisciredi* in relation to age and reproductive stage. *Amphibia–Reptilia* 12, 137–144.
- Macholán, N. (2006). A geometric morphometric analysis of the shape of the first upper molar in mice of the genus *Mus* (Muridae, Rodentia). *Journal of Zoology* 270, 672– 681.
- Mallow, D., Ludwig, D. & Nilson, G. (2003). True Vipers. Natural History and Toxinology of Old World Vipers. Malabar, Florida: Krieger Publishing Co.
- Marcus, L.F., Hingst-Zaher, E. & Zaher, H. (2000). Application of landmark morphometrics to skulls representing the orders of living mammals. *Hystrix* 11, 27–48.
- Nicola, P.A., Monteiro, L.R., Pessoa, L.M., Von Zuben, F.J., Rohlf, F.J. & Dos Reis, S.F. (2003). Congruence of hierarchical, localized variation in cranial shape and molecular phylogenetic structure in spiny rats, genus *Trinomys* (Rodentia: Echimyidae). <u>Biological Journal of</u> the Linnean Society 80, 385–396.
- Oxnard, C.E. (2000). Morphometrics of the primate skeleton and the functional and developmental underpinnings of species diversity. In *Development*, *Growth and Evolution*, 235–263. O'Higgins, P. & Cohn, M.J. (eds). London: Academic Press.
- Phisalix, M. (1968). La livrée des vipères de France. *Bulletin du Museum National d'Histoire Naturelle Paris* 4, 661–676.
- Pozio, E. (1980). Contributo alla sistematica di Vipera aspis

(L.) mediante analisi elettroforetica delle proteine contenute nel veleno. *Natura Milano* 71, 28–34.

- Rohlf, F.J. (1970). Adaptive hierarchical clustering schemes. *Systematic Zoology* 19, 58–82.
- Rohlf, F.J. (1998). On applications of geometric morphometrics to study of ontogeny and phylogeny. *Systematic Biology* 47, 147–158.
- Rohlf, F.J. (2005). *TPS–Series*. Stony Brook: Department of Ecology and Evolution, State University of New York.
- Rohlf, F.J. & Slice, D.E. (1990). Extensions of the Procrustes method for the optimal superimposition of landmarks. *Systematic Zoology* 39, 40–59.
- Rohlf, F.J. & Marcus, L.F. (1993). A revolution in morphometrics. <u>Trends in Ecology and Evolution 8, 129–</u> 132.
- Saint Girons, H. (1978). Morphologie externe comparée et systématique des vipères d'Europe (Reptilia, Viperidae). *Revue Suisse de Zoologie* 85, 565–595.
- Saint Girons, H. (1997). Vipera aspis (Linnaeus, 1758). In Atlas of Amphibians and Reptiles in Europe, 386–387.
 Gasc, J.P., Cabela, A., Crnobrnja-Isailovic, J., Dolmen, D., Grossenbacher, K., Haffner, P., Lescure, J., Martens, H., Martìnez Rica, J.P., Maurin, H., Oliveira, M.E., Sofianidou, T.S., Veith, M. & Zuiderwijk, A. (eds). Paris: Societas Europaea Herpetologica and Muséum National d'Histoire Naturelle (IEGP/SPN).

- Ursenbacher, S., Conelli, A., Golay, P., Monney, J.-C., Zuffi, M.A.L., Thiery, G., Durand, T. & Fumagalli, L. (2006). Phylogeography of the asp viper (*Vipera aspis*) inferred from mitochondrial DNA sequence data: evidence for multiple Mediterranean refugial areas. *Molecular Phylogenetics and Evolution* 38, 546–552.
- Zelditch, M.L., Swiderski, D.L., Sheets, H.D. & Fink, W.L., (2004). *Geometric Morphometrics for Biologists: a Primer*. San Diego: Elsevier Academic Press.
- Zuffi, M.A.L. (2002). A critique of the systematic position of the asp viper subspecies Vipera aspis aspis (Linnæus, 1758), Vipera aspis atra Meisner, 1820, Vipera aspis francisciredi Laurenti, 1768, Vipera aspis hugyi Schinz, 1833 and Vipera aspis zinnikeri Kramer, 1958. Amphibia–Reptilia 23, 191–213.
- Zuffi, M.A.L. & Bonnet, X. (1999). Italian subspecies of the asp viper, *Vipera aspis*: patterns of variability and distribution. *Italian Journal of Zoology* 66, 87–95.
- Zuffi, M.A.L., Gentilli, A., Razzetti, E. & Scali, S. (2003). Transition hybridization areas in parapatric *Vipera aspis* subspecies from northern Italy. *Biota* 3 (2002), 191– 196.

Accepted: 7 May 2009

					Catalogue	
Taxon	Sex ¹	Locality ²	Collection ³	Box^4	number	Structure
francisciredi	m	Valley of Sestaione (Lucca)	Calabresi	1	c96	frontal
francisciredi	m	Valley of Sestaione (Lucca)	Calabresi	2	c96	both
francisciredi	f	Vinara Aguasse (Lucca)	Calabresi	4	c60	frontal
francisciredi	m	Bagni di Lucca (Lucca)	Calabresi	7	c95	frontal
francisciredi	f	Bagni di Lucca (Lucca)	Calabresi	8	c95	both
francisciredi	m	San Rossore (Pisa)	Calabresi	16	1	both
francisciredi	f	San Rossore (Pisa)	Calabresi	17	2	both
francisciredi	m	Radda in Chianti (Florence)	Calabresi	23	c699	frontal
francisciredi	m	Treviso	Calabresi	3	c139	both
francisciredi	m	Udine	Calabresi	10	c399	frontal
francisciredi	f	Udine	Calabresi	12	c399	both
francisciredi	m	Tuscany	MSNT		B741	frontal
francisciredi	f	Tuscany	MSNT		B170	both
francisciredi	m	Tuscany	MSNT		B741/3	cranial base
francisciredi	m	Udine	Calabresi	6	c399	both
francisciredi	f	Udine	Calabresi	11	c399	both
francisciredi	f	Poggia Riparghera (Florence)	Calabresi	20		frontal
francisciredi	f	Valle Ombrosa (Florence)	Calabresi	22	c697	frontal
aspis	m	Les Moutiers (France)	MSNT		T65	both
aspis	m	Riva Valdobbia (Vercelli)	Calabresi	30	c490	both
aspis	f	Alpi d'Ossola (Verbania)	Calabresi	31	c132	both
aspis	f	Calasca Monte Rosa (Verbania)	Calabresi	34	c87	frontal
aspis	m	Monte Rosa	Calabresi	35	c315	frontal
aspis	f	Monte Rosa	Calabresi	36	c315	both
aspis	m	Valley of Lanzo (Turin)	Calabresi	37	c626	both
hugyi	f	Neto (Crotone)	Calabresi	2	c578	both
hugyi	m	Neto (Crotone)	Calabresi	3	c573	frontal

APPENDIX

Museum catalogue numbers and locality of collection for the specimens

					Catalogue	
Taxon	Sex ¹	Locality ²	Collection ³	Box^4	number	Structure
hugyi	m	Simmeri (Catanzaro)	Calabresi	5	c580	both
hugyi	m	Simmeri (Catanzaro)	Calabresi	8	c580	frontal
hugyi	m	Serra S. Bruno (Catanzaro)	Calabresi	12	c620	both
hugyi	m	Serra S. Bruno (Catanzaro)	Calabresi	13	c620	frontal
hugyi	m	Serra S. Bruno (Catanzaro)	Calabresi	15	c620	both
hugyi	m	Serra S. Bruno (Catanzaro)	Calabresi	16	c620	both
hugyi	m	Serra S. Bruno (Catanzaro)	Calabresi	22	c541	frontal
hugyi	m	Serra S. Bruno (Catanzaro)	Calabresi	23	c620	both
hugyi	f	Serra S. Bruno (Catanzaro)	Calabresi	26	c620	both
hugyi	f	Serra S. Bruno (Catanzaro)	Calabresi	27	c620	both
hugyi	f	Mongiana Calabra	Calabresi	32	c287	both
hugyi	m	Palermo	Calabresi	33	c640	cranial base
hugyi	f	Catania	Calabresi	36	c317	frontal
hugyi	m	Serra S. Bruno (Catanzaro)	Calabresi	14	c620	both
hugyi	f	Montecristo Island (Livorno)	Calabresi	3	c199	both
hugyi	m	Montecristo Island (Livorno)	Calabresi	4	c199	both
hugyi	f	Montecristo Island (Livorno)	Calabresi	5	c199	frontal
hugyi	m	Montecristo Island (Livorno)	Calabresi	6	c199	frontal
hugyi	m	Torre Loto (Taranto)	MSNT		1169	cranial base
hugyi	f	Montecristo Island (Livorno)	MSNT		1187	both
hugyi	f	Serra S. Bruno (Catanzaro)	Calabresi	24	c541	frontal
aspis*francisciredi	m	Cergnago Mortara (Pavia)	Calabresi	17	c213	both
aspis*francisciredi	m	Cergnago Mortara (Pavia)	Calabresi	21	c312	both
aspis*francisciredi	f	Cergnago Mortara (Pavia)	Calabresi	22	c312	both
aspis*francisciredi	f	Cergnago Mortara (Pavia)	Calabresi	23	c312	both
aspis*francisciredi	f	Cergnago Mortara (Pavia)	Calabresi	23	c312	cranial base
aspis*francisciredi	m	Cergnago Mortara (Pavia)	Calabresi	24	c312	frontal
aspis*francisciredi	m	Cergnago Mortara (Pavia)	Calabresi	25	c312	both
aspis*francisciredi	m	Cergnago Mortara (Pavia)	Calabresi	26	c312	both
francisciredi*hugyi	m	Valley of Clanio (Avellino)	MSNT		1172	both
berus	m	Morbegno (Sondrio)	Calabresi	F	c414	frontal
berus	f	Island of Ariano (Ferrara)	Calabresi	G	c530	both
berus	f	no locality	MSNT		701	cranial base
ammodytes	f	Krk Island	MSNT		1186	both

¹Sex: m, male; f, female.

²Locality (province).

³Collection: Calabresi – historical collection prepared by Enrica Calabresi in 1924 and presently housed at the 'La Specola' Zoological Museum of the University of Florence; MSNT – Museo di Storia Naturale e del Territorio of the University of Pisa.

⁴Box: reference number of specimens examined by Calabresi (1924).